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Key indicators

Single-crystal X-ray study T = 296 KMean $\sigma(\text{C}-\text{C}) = 0.004 \text{ Å}$ R factor = 0.035 wR factor = 0.087 Data-to-parameter ratio = 15.5

For details of how these key indicators were automatically derived from the article, see http://journals.iucr.org/e.

© 2005 International Union of Crystallography Printed in Great Britain – all rights reserved 1,2,3-Tri-O-acetyl-5-deoxy-5-methylthio- β -D-ribo-furanose

In the structure of the title compound, $C_{12}H_{18}O_7S$, no alteration of the relative configuration compared with D-(-)-ribose is observed.

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Comment

5-Methylthioribose (MTR) was synthesized from D-(-)-ribose following the procedure described by Ishikura *et al.* (1962). We report here the crystal structure of 1,2,3-tri-*O*-acetyl-5-deoxy-5-methylthio- β -D-ribofuranose, (I), obtained by acetylation of MTR. Acetylation of free hydroxyl groups is a common protecting procedure in carbohydrate chemistry (Greene & Wuts, 1999). Acetylated carbohydrates are extremely useful intermediates in the synthesis of natural products or analogues. Following a procedure devised by Euzen *et al.* (2005), the title compound was further converted into the ammonium salt of 5-deoxy-5-methylthio- β -D-ribofuranose 1-phosphate (MTR-1P). MTR-1P is known to be an intermediate in the methionine salvage pathway (Bumann *et al.*, 2004) and will therefore be utilized for enzymatic studies.



The crystal structure of (I) was determined in order to confirm its molecular structure. As expected, no alteration of the relative configuration compared with D-(-)-ribose is observed. The anomeric centre has the β configuration. The ring system shows a C_2' -exo conformation, with a pseudorotation phase angle of -24° (Saenger, 1984) and a C3-C4-O-C1 torsion angle of -3.5 (2)°. There are clear indications for an anomeric effect at C1: the 1'-O-acetyl group is oriented axially [O12-C1-O-C4 -88.8 (2)°], and C1-O is shorter by 0.019 Å and C1-O12 is longer by 0.023 Å than the standard bond lengths in furanoses of 1.421 (12) and 1.410 (14) Å, respectively (Allen *et al.*, 1992).

Experimental

5-Methylthioribose (570 mg, 3.1 mmol) was dissolved in dry pyridine (6.5 ml) under nitrogen and cooled to 273 K. Acetic anhydride (3 equivalents, 2.7 ml, 28.5 mmol) was added to the solution *via* a syringe over 5 min. The reaction mixture was stirred at room temperature for 18 h, and then the reaction was treated with icewater (40 ml) and stirred for 15 min. The mixture was then extracted with dichloromethane. The organic layer was washed with 1 M HCl

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 $(2 \times 50 \text{ ml})$ and water $(2 \times 50 \text{ ml})$, dried over MgSO₄ and concentrated in vacuo. Purification of the crude product by flash column chromatography on silica gel with cyclohexane-tert-butyl methyl ether (1:1) as solvent gave the title compound (300 mg, 31%). Crystals of (I) were obtained by evaporation of a cyclohexane/tert-butyl methyl ether (2:3) solution at room temperature. Analysis: $R_{\rm F}$ 0.32 (cyclohexane/*tert*-butyl methyl ether, 1:1, silica-gel plate 60 F_{254}); ¹H NMR (400 MHz, CDCl₃, δ, p.p.m.): 2.11, 2.08, 2.06 (3s, 9H, CH₃CO), 2.14 (s, 3H, CH₃S), 2.75 (d, 2H, J = 5.8 Hz, H5), 4.36 (q, 1H, J = 6.0 Hz, H4), 5.33 (m, 1H, H2), 5.36 (m, 1H, H3), 6.12 (bs, 1H, H1); ¹³C NMR (75 MHz, CDCl₃, *b*, p.p.m.): 16.6 (CH₃S), 20.6, 20.6, 21.2 (CH₃CO), 37.5 (C5), 73.0 (C3), 74.5 (C2), 81.3 (C4), 98.3 (C1), 169.2, 169.5, 169.8 (C=O); high-resolution MS (ESI+), m/z, calculated for C₁₂H₁₈O₇NaS: [M+Na]⁺ 329.0670; found: 329.0661; IR (solid, v, cm⁻¹): 3014, 2928, 1751, 1431, 1363, 1314, 1260, 1213, 1124, 1085, 1060, 1033, 1004.

Crystal data

C12H18O7S $M_r = 306.32$ Monoclinic, P2 a = 8.6831 (4) Å b = 9.8249 (5) Å c = 9.6824 (5) Å $\beta = 110.766 \ (2)^{\circ}$ $V = 772.35 (7) \text{ Å}^3$ Z = 2

 $D_{\rm r} = 1.317 {\rm Mg m}^{-3}$ Mo $K\alpha$ radiation Cell parameters from 1624 reflections $\theta = 2.3 - 22.2^{\circ}$ $\mu = 0.24 \text{ mm}^{-1}$ T = 296 (2) K Prism, colourless $0.43 \times 0.30 \times 0.16 \ \text{mm}$



Figure 1

The molecular structure of (I), showing the atomic numbering scheme and 50% probability displacement ellipsoids.

Data collection

188 parameters

Siemens SMART 1K CCD area- detector diffractometer ω scans Absorption correction: multi-scan (SADABS; Sheldrick, 1996)	2912 independent reflections 2272 reflections with $I > 2\sigma(I)$ $R_{int} = 0.015$ $\theta_{max} = 27.9^{\circ}$ $h = -11 \rightarrow 11$
$T_{\min} = 0.853, T_{\max} = 0.963$	$k = -12 \rightarrow 11$
Refinement	
Refinement on F^2	$w = 1/[\sigma^2(F_o^2) + (0.0424P)^2]$
$R[F^2 > 2\sigma(F^2)] = 0.035$	+ 0.0345P]
$wR(F^2) = 0.087$	where $P = (F_0^2 + 2F_c^2)/3$
S = 1.04	$(\Delta/\sigma)_{\rm max} = 0.008$
2012 reflections	$\Delta \rho = 0.11 \text{ e} \text{ Å}^{-3}$

 $\Delta \rho_{\rm max} =$ $\Delta \rho_{\rm min} = -0.14 \text{ e } \text{\AA}^{-3}$ Absolute structure: Flack (1983), with 1136 Friedel pairs Flack parameter: 0.04 (8)

Table 1 Selected geometric parameters (Å, °).

H-atom parameters constrained

C1-0	1.402 (3)	C2-C3	1.524 (3)
C1-O12	1.433 (3)	C3-C4	1.525 (3)
C1-C2	1.514 (3)	C4-O	1.450 (3)
0 - C4 - C3	105.86 (17)	C1 - C2 - C3	101.47 (18)
O-C1-O12	110.60 (18)	C2-C3-C4	103.60 (18)
O12-C1-C2	105.43 (19)	C1-O-C4	110.02 (16)
O12-C1-O-C4	-88.8 (2)	C1-C2-C3-C4	32.5 (2)
C4-O-C1-C2	25.0 (2)	C2-C3-C4-O	-19.0(2)
O-C1-C2-C3	-35.7 (2)	C3-C4-O-C1	-3.5 (2)

H atoms were placed in calculated positions (C-H = 0.97-0.98 Å) and were included in the refinement in a riding-model approximation, with $U_{iso}(H) = 1.2U_{eq}(C)$, or $1.5U_{eq}(C)$ for methyl H.

Data collection: SMART (Bruker, 2003); cell refinement: SAINT (Bruker, 2003); data reduction: SAINT; program(s) used to solve structure: SHELXTL (Sheldrick, 2003); program(s) used to refine structure: SHELXTL; molecular graphics: SHELXTL; software used to prepare material for publication: SHELXTL.

References

Allen, F. H., Kennard, O. Watson, D. G., Brammer, L., Orpen, A. G. & Tavlor, R. (1992). International Tables for Crystallography, Vol. C, edited by A. J. C. Wilson, pp. 685-706. Dordrecht: Kluwer Academic Publishers.

Bruker (2003). SMART (Version 5.059) and SAINT (Version 6.45a). Bruker AXS Inc., Madison, Wisconsin, USA.

Bumann, M., Djafarzadeh, S., Oberholzer, A. E., Bigler, P., Altmann, M., Trachsel, H. & Baumann, U. (2004). J. Biol. Chem. 279, 37087-37094.

Euzen, R., Ferrieres, V. & Plusquellec, D. (2005). J. Org. Chem. 70, 847-855. Flack, H. D. (1983). Acta Cryst. A39, 876-881.

Greene, T. W. & Wuts, P. G. M. (1999). Protective Groups in Organic Synthesis, p. 149. New York: Wiley & Sons Inc.

Ishikura, Y., Kanazawa, T. & Sato, T. (1962). Bull. Chem. Soc. Jpn, 35, 731-735.

Saenger, W. (1984). Principles of Nucleic Acid Structure, pp. 16-20. New York: Springer Verlag.

Sheldrick, G. M. (1996). SADABS. University of Göttingen, Germany.

Sheldrick, G. M. (2003). SHELXTL. DOS/Windows/NT Version 6.14. Bruker AXS Inc., Madison, Wisconsin, USA.